

IKK β KinEASE™ FP Fluorescein Green Assay

Catalog # 32-014

Sufficient reagents for two 384-well plates per kit.

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NOT RECOMMENDED OR INTENDED
FOR DIAGNOSIS OF DISEASE IN
HUMANS.**

DO NOT USE IN HUMANS.

I. STORAGE AND STABILITY

Storage: Upon receipt, store individual components at recommended temperatures. Store the 384-well plates at room temperature. Store all other components at -20°C.

Stability: Components stable for 6 months from date of shipment if stored and handled correctly. We recommend that all enzymes to be used with this kit are stored as aliquots and a fresh aliquot used for each experiment.

II. ASSAY OVERVIEW

In this assay, a phosphorylated peptide has been labeled with a green fluorescent dye. This phosphorylated tracer has a low molecular weight and thus a low fluorescence polarization value. The phosphorylated tracer binds to a phospho-specific antibody to form a high molecular weight complex with a high polarization value.

In a kinase reaction, a peptide or a protein substrate (non-fluorescently labeled) is phosphorylated by the kinase in the presence of ATP and Magnesium to form a phosphorylated product. This phosphorylated product competes with the tracer for binding to the phospho-specific antibody. As increasing amounts of phosphorylated product are formed from the kinase reaction, there is a reduction in the binding of tracer to antibody resulting in a decrease in the fluorescence polarization value.

IKK β KinEASE™ FP Fluorescein Green Assay supplied in this kit offers a method for assaying IKK β using preferred substrate and a generic detection system.



Related Product: IKK β , Catalog # 14-485

III. SYSTEM COMPONENTS

A. Provided Kit Components

Prior to use, each reagent should be vortexed, and then centrifuged to collect residual liquid trapped in the vial cap. **Please note the STK Tracer solution is light sensitive.**

Reagent	Catalog #	Stock	Volume Supplied	Storage
STK Substrate 1	12-548	10mM	4 x 120 μ l	-20°C
STK Antibody	35-002	20X	237 μ l	-20°C
STK Tracer – Fluorescein	20-287	100X	48 μ l	-20°C
10X KinEASE™ Buffer	20-302	10X	5ml	-20°C
10X KinEASE™ Enzyme Dilution Buffer	2004306	10X	1ml	-20°C
MgCl ₂	20-303	1M	500 μ l	-20°C
DTT	20-265	1M	450 μ l	-20°C
ATP	20-306	10mM	300 μ l	-20°C
EDTA	20-307	0.5M	2ml	-20°C
384 Well KinEASE™ Plate	30-014		2 plates	RT

Note: Individual buffer components should be stored at -20°C until ready to use

B. Recommended Buffers

Buffer	5X Buffer
Reaction Buffer	250mM HEPES pH 7.2, 0.05% BSA (Probumin™, Chemicon Catalog # 3225-80), 10mM MgCl ₂ , 5mM DTT (DTT to be added immediately prior to use)
Detection Buffer	250mM HEPES pH 7.2, 0.05% BSA (Probumin™, Chemicon Catalog # 3225-80), 5mM DTT (DTT to be added immediately prior to use)

Buffer	1X Buffer
Enzyme Dilution Buffer	50mM HEPES pH 7.2, 0.1% BSA (Probumin™, Chemicon Catalog # 1003512), 0.01% Brij-35, 0.1mM EDTA, 1mM DTT (DTT to be added immediately prior to use)

IV. ASSAY PROCEDURE

Safety Warnings and Precautions: The IKK β KinEASE™ FP Fluorescein Green Assay is designed for research use only. All chemicals should be considered potentially hazardous and principles of good laboratory practice should be followed.

A. Kinase Reaction Module

Prepare sufficient volume of each solution based on the number of assays to be performed, plus a slight overage to account for pipetting inaccuracies (either 10-20% extra or one extra assay point is generally sufficient). **Note:** A precipitate may be observed in the thawed 0.5M EDTA (Catalog # 20-307). Warm the tube to 37°C for 15 minutes and vortex to bring into solution.

Reaction Solution Preparation

- 5X Reaction Buffer:** For 1ml of 5X Reaction Buffer combine 485 μ l water, 500 μ l 10X KinEASE™ Buffer, 10 μ l 1M MgCl₂, and 5 μ l 1M DTT. The total amount of 5X Reaction Buffer required will depend upon the number of experimental assay points. The 1ml volume should be scaled up (or down) according to the particular requirements of the experiment.
- 1X Reaction Buffer (for use in the No Enzyme Wells, Buffer Control Wells and Tracer Control Wells):** Prepare the 1X Reaction Buffer by diluting the 5X Reaction Buffer 5-fold with water (e.g., for 1ml of 1X Reaction Buffer combine 800 μ l water and 200 μ l 5X Reaction Buffer).
- 1X Enzyme Dilution Buffer (EDB):** For 1ml of the 1X EDB combine 899 μ l water, 100 μ l of 10X KinEASE™ Enzyme Dilution Buffer, and 1 μ l of 1M DTT. The 1ml volume should be scaled up (or down) according to the particular requirements of the experiment.
- 2.5X ATP Working Solution:** Prepare a 250 μ M ATP Working Solution in 1.25X Reaction Buffer (e.g., for 1ml of the 2.5X ATP Working Solution combine 725 μ l water, 250 μ l 5X Reaction Buffer, and 25 μ l 10mM ATP). 10 μ l of the 2.5X ATP Working Solution is required per well.
- 2.5X STK Substrate 1 Working Solution:** Prepare a 500 μ M STK Substrate 1 Working Solution in 1.25X Reaction Buffer (e.g., for 1ml of STK Substrate 1 Working Solution combine 700 μ l water, 250 μ l 5X Reaction Buffer and 50 μ l of 10mM STK Substrate 1). 10 μ l of 2.5X STK Substrate 1 Working Solution is required per well.
- 5X IKK β Working Solution:** Prepare the IKK β Working Solution in 1X KinEASE™ EDB at a concentration of 5X the required final reaction concentration. 5 μ l of IKK β Working Solution is required per well.

Reaction Protocol

- Set up 3 assay controls as detailed below.

Buffer Control Wells (assay background)

25 μ l 1X Reaction Buffer

Tracer Control Wells (min. mP value)

25 μ l 1X Reaction Buffer

No Enzyme Wells (max. mP value)

10 μ l STK Substrate 1 Working Solution

5 μ l 1X Reaction Buffer

10 μ l ATP Working Solution

- Set up reactions as detailed below.

Plus Enzyme Wells

10 μ l STK Substrate 1 Working Solution
 5 μ l IKK β Working Solution
 10 μ l ATP Working Solution

The reactions are started by addition of the 10 μ l ATP Working Solution. When using the same concentration of enzyme (e.g. EC₇₀ value), the substrate working solution and enzyme working solutions can be combined and added as 15 μ l per well.

If required, 0.5 μ l DMSO may be added to a set of control wells to simulate compound addition.

- Incubate reactions for the required length of time at the appropriate temperature. To ensure that all reaction components have collected in the bottom of the wells, very gently tap the bottom of the plate against the bench top. **Note:** Optimum kinase reaction incubation time and temperature should be determined by the end user. Typical kinase reaction conditions used at Upstate are 30-60 minutes at room temperature with constant agitation.

B. Detection Module

Detection Solution Preparation

- 5X Detection Buffer:** Prepare the 5X Detection Buffer by diluting the 10X KinEASE™ Buffer 2-fold and adding DTT to a concentration 5mM (e.g. for 1ml of 5X Detection Buffer combine 495 μ l water, 500 μ l of 10X KinEASE™ Buffer and 5 μ l of 1M DTT).
- 1X Detection Buffer (for use in the Buffer Control Wells and Tracer Control Wells):** Prepare the 1X Detection Buffer by diluting the 5X Detection Buffer 5-fold with water (e.g., for 1ml of 1X Detection Buffer add 200 μ l 5X Detection Buffer to 800 μ l water).
- STK Stop Mix:** Prepare STK Stop Mix containing STK Tracer and EDTA in 1X Detection Buffer by diluting the STK Tracer 100-fold and adding EDTA to a concentration of 140mM (e.g., for 1ml of STK Stop Mix combine 510 μ l water, 280 μ l 0.5M EDTA pH 7.2, 200 μ l 5X Detection Buffer and 10 μ l 100X STK Tracer). 5 μ l of STK Stop Mix is required per well.
- STK Antibody Mix:** Prepare STK Antibody Mix containing STK Antibody in 1X Detection Buffer by diluting the STK Antibody 20-fold (e.g., for 1ml of STK Antibody Mix combine 750 μ l water, 200 μ l 5X Detection Buffer and 50 μ l 20X STK Antibody). 5 μ l of 5X STK Antibody Mix is required per well.

Detection Protocol

- Add the following to the 3 sets of assay controls wells.

Buffer Control Wells (assay background)

10 μ l 1X Detection Buffer

Tracer Control Wells (min. mP value)

5 μ l of STK Stop Mix
 5 μ l 1X Detection Buffer

No Enzyme Wells (max. mP value)

5 μ l STK Stop Mix
 5 μ l STK Antibody Mix

- Stop the reactions by adding 5 μ l/well STK Stop Mix.
- Add 5 μ l/well STK Antibody Mix. **Note:** Once the STK Stop Mix has been added to the Tracer Control wells, the STK Stop Mix and STK Antibody Mix may be combined and added as 10 μ l/well.

4. Incubate the plate for a minimum of 4 hours at room temperature (assay signal is stable for up to 24 hours).
5. Read plate on a Fluorescence Polarization Reader. The end user must optimize their instrument parameters for optimal assay sensitivity. Recommended parameters for the MDC Analyst AD used at Upstate are as follows:

Method: *Fluorescence Polarization*
Excitation: *485nm (bw20)*
Emission: *530nm (bw25)*
Mirror: *Dichroic 505*
Lamp: *Continuous*
Z-height: *3mm (to be determined for individual readers)*
Readings per well: *1*
Integration time: *100000 μ sec*

Attenuator: *out*
PMT setup: *Smartread Sensitivity 2*
Excitation Polarizer: *S (static)*
Emission Polarizer: *SP (dynamic)*
G Factor: *(to be determined for individual readers)*
Select Buffer Control Wells for background subtraction.

General Assay Notes

- It is recommended that the MgCl₂ final reaction concentration should not exceed 5mM in the reaction (*i.e.*, 25mM in the 5X Reaction Buffer). If a higher concentration of MgCl₂ is required then do not exceed 10mM MgCl₂ (*i.e.*, 50mM in the 5X Reaction Buffer) and run an enzyme reaction with stop mix and Antibody mix added prior to ATP addition to confirm that the reaction is stopped under these conditions.
- The 10X KinEASE™ Buffer (Catalog # 20-302) is used to prepare 5X working stocks to ensure a final 1X buffer concentration in the assay. These working buffers should be stored at 4°C until ready to use. If long-term storage (one week to six months) is required, DTT should be omitted until assay is to be performed and sodium azide added to the 5X buffer at a final concentration of 0.05%.

V. APPENDIX A - Assay Optimization

Determination of Enzyme Concentration for Screening

Using the optimized assay conditions, a suitable enzyme concentration for screening applications may then be determined by following the procedure outlined below.

Set up an enzyme titration with two-fold serial dilutions of enzyme and appropriate controls (No Enzyme Wells, Tracer Control Wells and Buffer Control Wells) according to the standard assay protocol.

Analyze the results obtained by plotting a graph of enzyme concentration (Units/ml) vs. Fluorescence Polarization (mP) (see below for example).

Determine the EC₇₀ mP using the following formula:

$$EC_{70} \text{ mP} = ((\text{No Enzyme Control} - \text{Tracer Control}) \times 0.3) + \text{Tracer Control}$$

From the graph of enzyme concentration (Units/ml) vs. Fluorescence Polarization (mP) determine the concentration of enzyme that correlates to the EC₇₀ mP value calculated above. This enzyme concentration (EC₇₀ Units/ml) is recommended for screening applications.

