

Protein A Agarose, Fast Flow
(10ml packed beads)
Catalog # 16-156
Lot # 20678

Product Description: Recombinant Protein A covalently coupled to highly cross-linked 6% agarose beads. Suitable for medium and low-pressure chromatography. Stable in all aqueous buffers used in Protein A chromatography. Recommended for flow rates from 30 to 400cm/hr.

Storage and Stability: Stable for 1 year at 4°C from date of shipment.

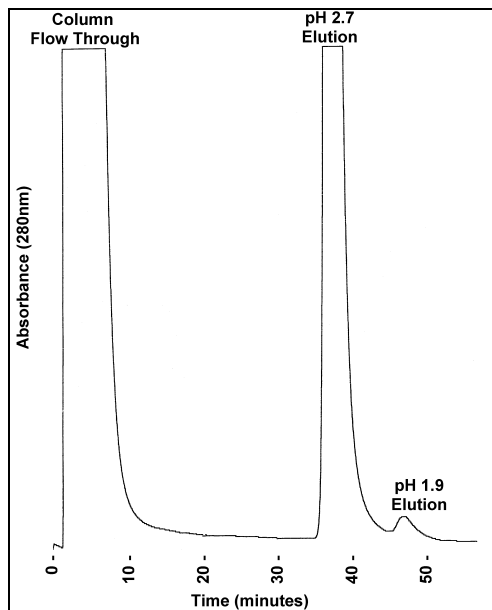
Physical Form: Liquid Suspension. Prior to use, wash the agarose beads with an appropriate buffer to remove the thimerosal.

Quantity and Formulation: 10ml packed beads, containing 3.5mg/ml recombinant Protein A. Suspended as a 50% slurry in phosphate-buffered saline containing 0.01% thimerosal for a final volume of 20ml. Binding capacity of 37mg human IgG/ml agarose.

FOR RESEARCH USE ONLY
NOT FOR USE IN HUMANS

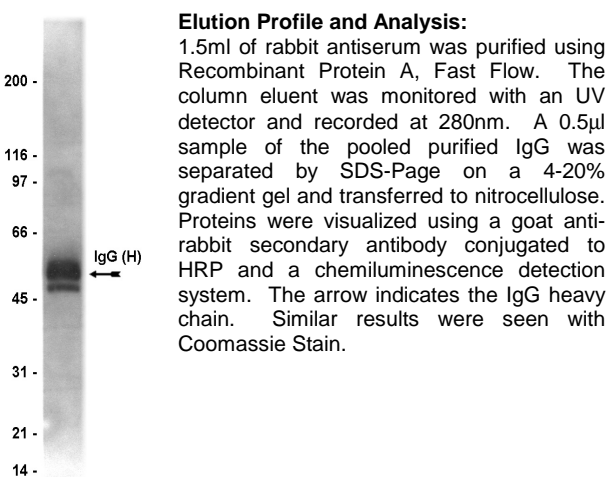
Quality Control Testing

Antibody Purification: Representative elution profile from a previous lot using a 1.7ml column of recombinant protein A agarose, fast flow, and the IgG from a 1.5ml sample of rabbit serum. The IgG was purified using a Waters™ 650E Advanced Protein Purification System.



Additional Research Applications

Immunoprecipitation: Use 100µl (50µl packed beads) to capture the immunoprecipitation immunocomplex.



Elution Profile and Analysis:

1.5ml of rabbit antiserum was purified using Recombinant Protein A, Fast Flow. The column eluent was monitored with a UV detector and recorded at 280nm. A 0.5µl sample of the pooled purified IgG was separated by SDS-Page on a 4-20% gradient gel and transferred to nitrocellulose. Proteins were visualized using a goat anti-rabbit secondary antibody conjugated to HRP and a chemiluminescence detection system. The arrow indicates the IgG heavy chain. Similar results were seen with Coomassie Stain.

Immunoprecipitation Protocol

1. Dilute the cell lysate before beginning the immunoprecipitation to roughly 1 μ g/ μ l total cell protein in a microcentrifuge tube with PBS.
2. Add antibody of choice to 500 μ g-1mg cell lysate.
3. Gently rock the reaction mixture at 4°C overnight.
4. Capture the immunocomplex by adding 100 μ l of washed **recombinant protein A agarose** bead slurry (50 μ l packed beads).
5. Gently rock the reaction mixture at 4°C for 2 hours.
6. Collect the agarose beads by pulsing (5 seconds in the microcentrifuge at 14,000 x g), and drain off the supernatant. Wash the beads 3 times with either ice-cold cell lysis buffer or PBS.
7. Resuspend the agarose beads in 50 μ l 2X Laemmli sample buffer.
8. The agarose beads can either be frozen for later use or suspended in Laemmli sample buffer and boiled for 5 minutes. Collect the beads by a microcentrifuge pulse. SDS-PAGE and subsequent immunoblot analysis can be performed on a sample of the supernatant.

Antibody Purification Protocol

1. Use a 0.45 μ filter to filter all buffers.
2. Degas all buffers prior to starting purification.
3. Pack **recombinant protein A agarose** into the column of choice, following the column manufacturer's instructions.
4. Wash and equilibrate packed protein A agarose with 10-20 column volumes of Tris buffered saline, pH 7.4 (TBS) at 1ml/minute flow rate.
5. Load sample containing IgG of interest onto column.
6. Wash column with 10-20 column volumes of TBS to remove unbound and non-specifically bound proteins. This can be determined by measuring the absorbance at 280nm with an inline UV detector. Alternatively, a post column sample can be collected and checked for protein.
7. Elute the bound IgG with 10ml of 50mM glycine pH 2.7, collecting 1ml fractions in tubes containing a antibody neutralization buffer (1M Tris, pH 8.0, 1.5M NaCl, 1mM EDTA). Eluted IgG can be determined by monitoring the absorbance at 280nm. **NOTE:** Some antibodies bind with high affinity to Protein A and will not elute at pH 2.7, in these cases, 50mM Glycine pH 1.9 may be used.
8. Wash the column with 10-20 volumes of TBS to restore the agarose column to neutral pH.
9. Store the column in TBS containing either 0.02% sodium azide or 0.01% thimerosal.