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### Magna ChIP™ G or EZ-Magna ChIP™ G Chromatin Immunoprecipitation Kits

Catalog # 17-611 or 17-409

MAGNA0002 Lot#DAM1469756

MAGNA0003 (Any Lot), MAGNA0005 (Any Lot)

Magna ChIP™ G & EZ-Magna ChIP™ G Kit Configurations	
Magna ChIP™ G (Cat. # 17-611)	EZ-Magna ChIP™ G (Cat. # 17-409)
MAGNA0002 (Store at 4°C)	MAGNA0002 (Store at 4°C)
MAGNA0003 (Store at -20°C)	MAGNA0005 (Store at -20°C)

#### Kit Components

MAGNA0002 Contents (17-611 or 17-409)		
Store at 4°C		
Component	Catalogue #	Quantity
Magnetic Protein G beads	CS200638	450 µl
ChIP Dilution Buffer	CS200624	12.5 ml
Low Salt Wash Buffer	CS200625	12.5 ml
High Salt Wash Buffer	CS200626	12.5 ml
LiCl Wash Buffer	CS200627	12.5 ml
TE Buffer	CS200628	12.5 ml
Cell Lysis Buffer	CS200634	5 ml
Nuclear Lysis Buffer	CS200623	5 ml
ChIP Elution buffer (w/o Proteinase K)	CS200629	5 ml
10X Glycine	20-282	11 ml
10X PBS	20-281	24 ml
Store the Following at Room Temperature Upon Receipt		
Spin Filters	20-290	22 Filters
Collection Tubes	20-291	22 Tubes
Bind Reagent A	20-292	25 ml
Wash Reagent B	20-293	12.5 ml
Elution Reagent C	20-294	1.5 ml
MAGNA0003 Contents (17-611)		
Store at -20°C		
Protease Inhibitor Cocktail II, 200X	20-283	110 µl <b>**Contains DMSO</b>
Proteinase K	20-298	600 µg in 60 µl
MAGNA0005 Contents (17-409)		
Store at -20°C		
Protease Inhibitor Cocktail II, 200X	20-283	110 µl <b>**Contains DMSO</b>
Proteinase K	20-298	600 µg in 60 µl
Control Primers	22-004	75 µL
Anti-RNA Polymerase II, clone CTD4H8	05-623B	25 µg
Normal Mouse IgG	12-371B	25 µg

**FOR RESEARCH USE ONLY  
DO NOT USE IN HUMANS**

**Product Description:** The Magna ChIP™ kits contain reagents optimized for immunoprecipitation of chromatin from mammalian cells. The EZ-Magna ChIP™ kits contain all elements of the Magna ChIP™ kits, but also include IP and PCR controls to ensure successful optimization of the assay.

For all Magna ChIP™ kits, detection of the DNA region, gene or promoter of interest in immunoprecipitated chromatin must be empirically determined by the researcher. PCR using promoter-specific primers is recommended for detection and analysis of enriched DNA.

For **EZ-Magna ChIP™ G** (Cat.# 17-409), the positive control antibody is a mouse monoclonal antibody that detects RNA Pol II of human, mouse, rat and yeast origins. The negative control is Normal Mouse IgG which controls for the non-specific immunoselection of chromatin by immunoglobulins.

The Control Primer mix is included for detection of a 166 base pair region of the human GAPDH promoter by both end-point and real-time quantitative PCR. Use of these primers with DNA from species other than human is not recommended.

**Quantity:** Two boxes containing all necessary reagents to perform 22 chromatin immunoprecipitation (ChIP) assays. [Supplied buffers are sufficient to generate chromatin from up to five 15 cm plates of cultured cells, each plate providing up to 10 chromatin preparations \(varies with cell and assay type\).](#)

**Storage and Stability:** Upon receipt, store components at the temperatures indicated on the labels. Kit components are stable for 1 year from date of shipment when stored as directed.

**Quality Control & Application Testing**

End Point and qPCR Analysis of Chromatin Immunoprecipitation:

Chromatin immunoprecipitation was performed using chromatin from HeLa cells and either anti-RNA Polymerase II (Cat.# 05-693B) or Normal Mouse IgG (Cat.# 12-371B) as the immunoprecipitating antibody. Purified DNA was then analyzed by end point PCR (Fig. 1, representative lot) or qPCR (Fig. 2, current lot) using Control Primers specific for the GAPDH promoter. Fold Enrichment reflects the ratio of RNA Pol II signal to that of IgG signals derived from a standard curve of Input DNA in qPCR.

Fig. 2

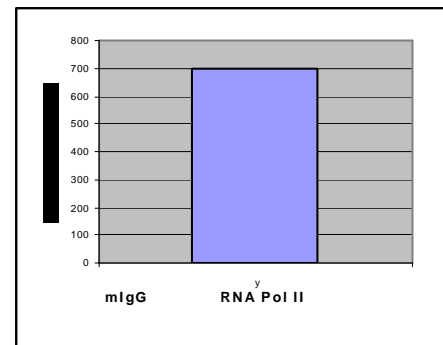


Fig. 1



Fig.1 Samples:

- Lane 1: No template control
- Lane 2: Mouse IgG IP
- Lane 3: Anti-RNA Polymerase II IP
- Lane 4: 2% chromatin Input

**General References:**

1. Das, Partha M, *et al* (2004). Chromatin immunoprecipitation assay. *BioTechniques* **37**: 961-9.
2. Luo, R X, *et al* (1998). Rb interacts with histone deacetylase to repress transcription. *Cell* **92**: 463-73.
3. Braunstein, M, *et al* (1996). Efficient transcriptional silencing in *Saccharomyces cerevisiae* requires a heterochromatin histone acetylation pattern. *Mol Cell Biol* **16**: 4349-56.
4. Buck, Michael J and Lieb, Jason D (2004). ChIP-chip: considerations for the design, analysis, and application of genome-wide chromatin immunoprecipitation experiments. *Genomics* **83**: 349-60.
5. Bernstein, Bradley E, *et al* (2004). The use of chromatin immunoprecipitation assays in genome-wide analyses of histone modifications. *Meth Enzymol* **376**: 349-60.