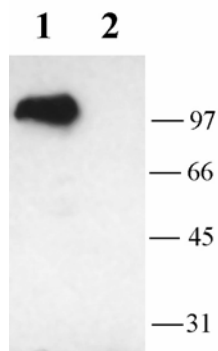


**RABBIT ANTI-Kv3.1b, VOLTAGE-GATED POTASSIUM CHANNEL  
AFFINITY PURIFIED  
POLYCLONAL ANTIBODY**

<b>CATALOG NUMBER:</b>	AB5188
<b>LOT NUMBER:</b>	
<b>QUANTITY:</b>	200 $\mu$ L
<b>CONCENTRATION:</b>	0.3 mg/mL (after reconstitution)
<b>SPECIFICITY:</b>	Recognizes a full length Kv3.1b protein. Does not cross react with any other potassium channel antigens tested so far.
<b>IMMUNOGEN:</b>	Purified peptide from rat Kv3.1b (amino acids 567-585) (Accession P25122).
<b>APPLICATIONS:</b>	Western blot: 1:200 using ECL on rat brain membranes. Immunohistochemistry on rat brain sections. Immunoprecipitation Dilutions should be made using a carrier protein such as BSA (1-3%) Optimal working dilutions must be determined by the end user.
<b>CONTROL ANTIGEN:</b>	Included free of charge with the antibody is 40 $\mu$ g of control antigen. The stock solution of the antigen can be made up using 100 $\mu$ L of sterile deionized water. For negative control, preincubate 1 $\mu$ g of peptide with 1 $\mu$ g of antibody for one hour at room temperature. Optimal concentrations must be determined by the end user.
<b>SPECIES REACTIVITIES:</b>	Rat, mouse, canine and <i>Macaca</i> . Other species have not been tested.
<b>FORMAT:</b>	Affinity purified immunoglobulin.
<b>PRESENTATION:</b>	Lyophilized from phosphate buffered saline, pH 7.4, containing 1% BSA, 5% sucrose as a stabilizer and 0.025% sodium azide as a preservative. Reconstitute with 200 $\mu$ L of sterile deionized water. Centrifuge antibody preparation before use (10,000 xg for 5 min).
<b>STORAGE/HANDLING:</b>	Maintain lyophilized material at -20°C for up to 12 months after date of receipt. After reconstitution maintain at -20°C in undiluted aliquots for up to 6 months. Avoid repeated freeze/thaw cycles.
<b>REFERENCE:</b>	Elezgarai, I., et al., <i>Neuroscience</i> (2003) <b>118</b> :889-898.

## SUGGESTED WESTERN BLOT PROTOCOL

1. Mix the samples with sample-buffer X 2, and heat 10 min at 70°C.
2. 5-50  $\mu$ L applied to Minigel lane (0.75-1.5 mm width) and run at standard conditions. (60 mA for 2 1.5 mm Minigel gels, 1.4 h)
3. Transfer in semi-dry system under standard conditions (3 h 200 mA for two minigel gels)
4. Stain the transferred bands with Chemicon BLOT-*FastStain* (Catalog Number 2076).
5. Destain with deionized water.
6. Block with 5% non-fat milk (Marvel or Carnation) in PBS, and 0.025 % sodium azide, overnight at 4°C. The non-fat milk should be dissolved freshly, centrifuged 10,000 rpm for 10 min, and filtered through glass filter (Gelman Acrodisc).
7. Incubation with first antibody 2 h at room temperature or overnight at 4°C in blocking solution. The antibody preparation should be centrifuged before use (10,000 g 5 min.). Optimal working dilutions and incubation time will need to be determined by the end user.
8. Wash 4 x 15 min. with PBS-0.1% tween 20. From this stage, azide should be omitted.
9. Incubation with the secondary antibody (HRP-conjugated goat anti-rabbit antibody, for example Chemicon Catalog Number AP132P, diluted appropriately) 1 h at room temperature.
10. Wash 4 x 15 min. with PBS-0.1% tween 20.
11. Perform ECL with commercial kits (*ChemLUCENT*, Chemicon Catalog Number 2600).



Western blotting of rat brain membranes (25  $\mu$ g/lane)

1. AB5188 (1:200)
2. AB5188 preincubated with control antigen peptide

**PROTOCOL FOR IMMUNOHISTOCHEMISTRY**Sacrifice and tissue processing

Rats are deeply anesthetized with pentobarbital sodium (Pental). Brain is fixed by transcardial perfusion, first with 50 mL of phosphate buffered saline (0.02M PBS, pH 7.4) containing heparin (5U/ml), then with 220 mL of ice-cold 4% paraformaldehyde in 0.1M PBS, pH 7.5 containing sucrose 4%. Brain is cut in coronal blocks and further fixed by immersion in the same fixative, refrigerated, for 1-2 hours. Brain blocks are transferred to 10% sucrose in 0.1M PBS and sectioned in a cryostat within 3 days. Brain sections, 30 µm thick, are floated in 0.1M PBS and then preserved in a cryopreservation buffer at -20°C. The cryopreservation buffer contains 40% ethylene glycol and 1% polyvinylpyrrolidone in 0.1M potassium acetate buffer, pH 6.5.

Staining procedure

1. Floating sections are rinsed in 0.02M PBS, 2x 5 minutes.
2. Endogenous peroxidase activity is quenched by incubation with 0.2 % hydrogen peroxide in 0.1M phosphate buffer pH 7.3 containing 0.2% Triton X-100 for 25 minutes at room temperature.
3. Sections are rinsed in 0.02M PBS, 2x 5 minutes.
4. Sections are incubated with the primary antiserum in a medium containing 0.3% Triton X-100, 0.05% Tween 20, 4% normal donkey serum (NDS), for 1 hour at room temperature and then overnight refrigerated.
5. Sections are rinsed in 0.02M PBS, containing 4% NDS, 2x 5 minutes.

Sections are incubated with biotinylated donkey anti-rabbit (from Chemicon USA, catalog number AP182B) diluted 1:400 in 0.02M PBS, containing 0.3% Triton X-100, 0.05% Tween 20, and 4% NDS, for 1 hour at room temperature and then overnight refrigerated.

1. Sections are rinsed in 0.02M PBS containing 4% NDS, 2x 5 min.
2. Sections are incubated with extravidin-peroxidase (Sigma Catalog number E2886) diluted 1:100 in 0.02M PBS, for 45 minutes at room temperature.
3. Sections are rinsed in 0.02M PBS, 3x 5 minutes.

Color development

1. Sections are incubated with a solution of diaminobenzidine at the concentration of 0.0125% and containing 0.05% nickel ammonium sulfate for 10 minutes at room temperature.
2. Sections are transferred to the same DAB solution but with added hydrogen peroxide at a final concentration of 0.0015%. Duration of incubation should be adjusted by the end user.
3. Sections are rinsed in 0.02M PBS, 4x 10 minutes.
4. Sections are mounted on glass slides (gelatinized or coated by any other type of adhesive material) and allowed to dry.
5. Sections are dehydrated in ascending series of ethanol concentrations (70%, 90%, 100%, 5 minutes in each), delipidated in xylene (10 minutes) and coverslipped in Permount (or any other xylene diluted adhesive).

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